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Project-Team MAGNOME

Models and Algorithms for the Genome

IN COLLABORATION WITH: Laboratoire Bordelais de Recherche en Informatique (LaBRI)

RESEARCH CENTER Bordeaux - Sud-Ouest

THEME Computational Biology and Bioinformatics

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Project-Team MAGNOME

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MAGNOME is a joint projet-team with the PRES Bordeaux (Universities Bordeaux 1 and Bordeaux Ségalen) and the CNRS (LaBRI UMR 5800). All of the members of MAGNOME are also members of the LaBRI.

1. Members

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2. Overall Objectives

2.1. Overall Objectives

One of the key challenges in the study of biological systems is understanding how the static information recorded in the genome is interpreted to become dynamic systems of cooperating and competing biomolecules. MAGNOME addresses this challenge through the development of informatic techniques for multi-scale modeling and large-scale comparative genomics:

- logical and object models for knowledge representation
- stochastic hierarchical models for behavior of complex systems, formal methods
- algorithms for sequence analysis, and
- data mining and classification.

We use genome-scale comparisons of eukaryotic organisms to build modular and hierarchical hybrid models of cell behavior that are studied using multi-scale stochastic simulation and formal methods. Our research program builds on our experience in comparative genomics, modeling of protein interaction networks, and formal methods for multi-scale modeling of complex systems.

New high-throughput technologies for DNA sequencing have radically reduced the cost of acquiring genome and transcriptome data, and introduced new strategies for whole genome sequencing. The result has been an increase in data volumes of several orders of magnitude, as well has a greatly increased density of genome sequences within phylogenetically constrained groups of species. MAGNOME develops efficient techniques for dealing with these increased data volumes, and the combinatorial challenges of dense multi-genome comparison.

2.2. Highlights

With clinical and academic partners MAGNOME participated in the development of a new rapid diagnostic test for yeast pathogens in the *Nakaseomycetes* class, based on a comparative annotation of six genomes[13].

These *de novo* 6 genomes – 5 genomes in the class *Nakaseomycetes*, 1 strain of genome of *S.cerevisiae* – were automatically annotate from their raw sequences using our YAGA software.

Through a long-standing collaboration between the LaBRI and Prof. Aline Lonvaud at the Institute of Vine and Wine Sciences of Bordeaux, and under the auspices of the ANR DIVOENI contract (2008-2012), we successfully completed the first comparative exploration of Oenococcus oeni pan-transcriptom code. The guidelines delivered partially lift the veil on how the genome of this lactic acid bacterium involved in wine fermentation globally adapts to its environment at a functional and an organisational level[16].

We released the first whole-genome metabolic model of the oleaginous yeast *Yarrowia lipolytica*, developed using our Pathtastic software and curated in collaboration with colleages from the INRA Grignon (model in MODEL1111190000 in Biomodels.net).

The complete implementation of the BioRica modeling framework was deposited with the APP and has been released [biorica]. BioRica was developed as an Inria Technology Development Action.

3. Scientific Foundations

3.1. Overview

Fundamental questions in the life sciences can now be addressed at an unprecedented scale through the combination of high-throughput experimental techniques and advanced computational methods from the computer sciences. The new field of *computational biology* or *bioinformatics* has grown around intense collaboration between biologists and computer scientists working towards understanding living organisms as *systems*. One of the key challenges in this study of systems biology is understanding how the static information recorded in the genome is interpreted to become dynamic systems of cooperating and competing biomolecules.

MAGNOME addresses this challenge through the development of informatic techniques for understanding the structure and history of eukaryote genomes: algorithms for genome analysis, data models for knowledge representation, stochastic hierarchical models for behavior of complex systems, and data mining and classification. Our work is in methods and algorithms for:

- Genome annotation for complete genomes, performing *syntactic* analyses to identify genes, and *semantic* analyses to map biological meaning to groups of genes [5], [9], [10], [50], [51].
- Integration of heterogenous data, to build complete knowledge bases for storing and mining information from various sources, and for unambiguously exchanging this information between knowledge bases [1], [3], [37], [38], [27].

- Ancestor reconstruction using optimization techniques, to provide plausible scenarios of the history of genome evolution [10], [7], [40], [56].
- Classification and logical inference, to reliably identify similarities between groups of genetic elements, and infer rules through deduction and induction [8], [6], [9].
- **Hierarchical and comparative modeling**, to build mathematical models of the behavior of complex biological systems, in particular through combination, reutilization, and specialization of existing continuous and discrete models [36], [25], [54], [30], [53].

The hundred- to thousand-fold decrease in sequencing costs seen in the past few years presents significant challenges for data management and large-scale data mining. MAGNOME's methods specifically address "scaling out," where resources are added by installing additional computation nodes, rather than by adding more resources to existing hardware. Scaling out adds capacity and redundancy to the resource, and thus fault tolerance, by enforcing data redundancy between nodes, and by reassigning computations to existing nodes as needed.

3.2. Comparative genomics

The central dogma of evolutionary biology postulates that contemporary genomes evolved from a common ancestral genome, but the large scale study of their evolutionary relationships is frustrated by the unavailability of these ancestral organisms that have long disappeared. However, this common inheritance allows us to discover these relationships through *comparison*, to identify those traits that are common and those that are novel inventions since the divergence of different lineages.

We develop efficient methodologies and software for associating biological information with complete genome sequences, in the particular case where several phylogenetically-related eukaryote genomes are studied simultaneously.

The methods designed by MAGNOME for comparative genome annotation, structured genome comparison, and construction of integrated models are applied on a large scale to:

- eukaryotes from the hemiascomycete class of yeasts [50], [51], [5], [9], [2], [10] and to
- prokaryotes from the lactic bacteria used in winemaking [28], [33], [26].

3.3. Comparative modeling

A general goal of systems biology is to acquire a detailed quantitative understanding of the dynamics of living systems. Different formalisms and simulation techniques are currently used to construct numerical representations of biological systems, and a recurring challenge is that hand-tuned, accurate models tend to be so focused in scope that it is difficult to repurpose them. We claim that, instead of modeling individual processes *de novo*, a sustainable effort in building efficient behavioral models must proceed incrementally. Hierarchical modeling is one way of combining specific models into networks. Effective use of hierarchical models requires both formal definition of the semantics of such composition, and efficient simulation tools for exploring the large space of complex behaviors. We have combined uses theoretical results from formal methods and practical considerations from modeling applications to define BioRica[19] [36], [54], a framework in which discrete and continuous models can communicate with a clear semantics. Hierarchical models in BioRica can be assembled from existing models, and translated into their execution semantics and then simulated at multiple resolutions through multi-scale stochastic simulation. BioRica models are compiled into a discrete event formalism capable of capturing discrete, continuous, stochastic, non deterministic and timed behaviors in an integrated and non-ambiguous way. Our long-term goal to develop a methodology in which we can assemble a model for a species of interest using a library of reusable models and a organismlevel "schematic" determined by comparative genomics.

Comparative modeling is also a matter of reconciling experimental data with models [4] [25] and inferring new models through a combination of comparative genomics and successive refinement [45], [46].

4. Application Domains

4.1. Function and history of yeast genomes

Yeasts provide an ideal subject matter for the study of eukaryotic microorganisms. From an experimental standpoint, the yeast *Saccharomyces cerevisiae* is a model organism amenable to laboratory use and very widely exploited, resulting in an astonishing array of experimental results. From a genomic standpoint, yeasts from the hemiascomycete class provide a unique tool for studying eukaryotic genome evolution on a large scale. With their relatively small and compact genomes, yeasts offer a unique opportunity to explore eukaryotic genome evolution by comparative analysis of several species.

- Yeasts are widely used as cell factories, for the production of beer, wine and bread and more recently of various metabolic products such as vitamins, ethanol, citric acid, lipids, etc.
- Yeasts can assimilate hydrocarbons (genera *Candida, Yarrowia* and *Debaryomyces*), depolymerise tannin extracts (*Zygosaccharomyces rouxii*) and produce hormones and vaccines in industrial quantities through heterologous gene expression.
- Several yeast species are pathogenic for humans, especially *Candida albicans*, *Candida glabrata*, *Candida tropicalis* and the Basidiomycete *Cryptococcus neoformans*.

The hemiascomycetous yeasts represent a homogeneous phylogenetic group of eukaryotes with a relatively large diversity at the physiological and ecological levels. Comparative genomic studies within this group have proved very informative [29], [42], [41], [32], [44], [2], [5].

MAGNOME applies its methods for comparative genomics and knowledge engineering to the yeasts through the ten-year old *Génolevures* program (GDR 2354 CNRS), devoted to large-scale comparisons of yeast genomes with the aim of addressing basic questions of molecular evolution. We developed the software tools used by the CNRS's genolevures.org web site. MAGNOME's MAGUS system for simultaneous genome annotation combines semi-supervised classification and rule-based inference in a collaborative web-based system that explicitly uses comparative genomics to simultaneously analyse groups of related genomes.

4.2. Alternative fuels and bioconversion

Oleaginous yeasts are capable of synthesizing lipids from different substrates other than glucose, and current research is attempting to understand this conversions with the goal of optimizing their throughput, production and quality. From a genomic standpoint the objective is to characterize genes involved in the biosynthesis of precursor molecules which will be transformed into fuels, which are thus not derived from petroleum. Biological experimentation by partner laboratories study lipid accumulation the oleaginous yeasts such as *Yarrowia lipolytica* starting from:

- pentoses, produced from lignin cellulose agricultural substrates following a biorefining strategy,
- glycerol, a secondary output of chemical production of biodiesel, and
- industrial residues.

Lipases from *Y. lipolytica* are of particular interest (see [34] for review). Experimental characterization of the lipid bodies produced from these substrates will aid in the identification of target genes which may serve for genetic engineering. This in turn requires the development of molecular tools for this class of yeasts with strong industrial potential. MAGNOME's focus is in acquiring genome sequences, predicting genes using models learned from genome comparison and sequencing of cDNA transcripts, and comparative annotation. Our overall goal is to define dynamic models that can be used to predict the behavior of modified strains and thus drive selection and genetic engineering.

4.3. Winemaking and improved strain selection

Yeasts and bacteria are essential for the winemaking process, and selection of strains based both on their efficiency and on the influence on the quality of wine is a subject of significant effort in the Aquitaine region. Unlike the species studied above, yeast and bacterial starters for winemaking cannot be genetically modified. In order to propose improved and more specialized starters, industrial producers use breeding and selection strategies.

Yeast starters from the *Saccharomyces* genus are used for primary, alcohol fermentation. Recent advances have made it possible to identify the genetic causes of the different technological differences between strains [49], [48], [47]. Manipulating the genetic causes rather than the industrial consequences is far more amenable to experimental development. An essential tool in identifying these genetic causes is comparative genomics.

Bacterial starters based on *Oenococcus oeni* are used in secondary, malolactic fermentation. Genetically, *O. oeni* presents a surprising level of intra-specific diversity, and clues that it may evolve more rapidly than expected. Studying the diversity of the O. oeni genomes has led to genetic tools that can be used to evaluate the predisposition of different strains to respond to oenological stresses. While identifying particular genes has been the leading strategy up to now, recently a new strategy based on comparative genomics has been undertaken to understand the impact and mechanisms of genetic diversity [28], [33], [26].

Starting from historical collaborations by Pascal Durrens and Elisabeth Bon with partners from the Institute for Wine and Vine Sciences in Bordeaux (ISVV), we have built an effective partnership between MAGNOME, the UMR Œnology–ISVV, and local industry, to apply our tools to large-scale comparative genomics of yeast and bacterial starters in winemaking.

4.4. Knowledge bases for molecular tools

Affinity binders are molecular tools for recognizing protein targets, that play a fundamental in proteomics and clinical diagnostics. Large catalogs of binders from competing technologies (antibodies, DNA/RNA aptamers, artificial scaffolds, etc.) and Europe has set itself the ambitious goal of establishing a comprehensive, characterized and standardized collection of specific binders directed against all individual human proteins, including variant forms and modifications. Despite the central importance of binders, they presently cover only a very small fraction of the proteome, and even though there are many antibodies against some targets (for example, >900 antibodies against p53), there are none against the vast majority of proteins. Moreover, widely accepted standards for binder characterization are virtually nonexistent.

Alongside the technical challenges in producing a comprehensive binder resource are significant logistical challenges, related to the variety of producers and the lack of reliable quality control mechanisms. As part of the ProteomeBinders and Affinomics projects, MAGNOME works to develop knowledge engineering techniques for storing, exploring, and exchanging experimental data used in affinity binder characterization. This work involves databases and tools for molecular interaction data [21] [39], standards for data exchange between peers [38], [43], [37] and reporting standards [3] [52].

5. Software

5.1. Inria Bioscience Resources

Participants: Olivier Collin [correspondant], Frédéric Cazals, Mireille Régnier, Marie-France Sagot, Hélène Touzet, Hidde de Jong, David Sherman, Marie-Dominique Devignes, Dominique Lavenier.

Inria Bioscience Resources is a portal designed to improve the visibility of bioinformatics tools and resources developed by Inria teams. This portal will help the community of biologists and bioinformatians understand the variety of bioinformatics projects in Inria, test the different applications, and contact project-teams. Eight project-teams participate in the development of this portal. Inria Bioscience Resources is developed in an Inria Technology Development Action (ADT).

5.2. Magus: Collaborative Genome Annotation

Participants: David James Sherman [correspondant], Pascal Durrens, Natalia Golenetskaya, Florian Lajus, Tiphaine Martin.

As part of our contribution the Génolevures Consortium, we have developed over the past few years an efficient set of tools for web-based collaborative annotation of eukaryote genomes. The MAGUS genome annotation system integrates genome sequences and sequences features, *in silico* analyses, and views of external data resources into a familiar user interface requiring only a Web navigator. MAGUS implements the annotation workflows and enforces curation standards to guarantee consistency and integrity. As a novel feature the system provides a workflow for *simultaneous annotation* of related genomes through the use of protein families identified by *in silico* analyses; this has resulted in a three-fold increase in curation speed, compared to one-at-a-time curation of individual genes. This allows us to maintain Génolevures standards of high-quality manual annotation while efficiently using the time of our volunteer curators.

MAGUS is built on: a standard sequence feature database, the Stein lab generic genome browser [55], various biomedical ontologies (http://obo.sf.net), and a web interface implementing a representational state transfer (REST) architecture [35].

For more information see magus.gforge.inria.fr, the MAGUS Gforge web site. MAGUS is developed in an Inria Technology Development Action (ADT).

5.3. YAGA: Yeast Genome Annotation

Participants: Pascal Durrens, Tiphaine Martin [correspondant].

With the arrival of new generations of sequencers, laboratories, at a lower cost, can be sequenced groups of genomes. You can no longer manually annotate these genomes. The YAGA software's objective is to syntactically annotate a raw sequence (genetic element: gene, CDS, tRNA, centromere, gap, ...) and functionally as well as generate EMBL files for publication. The annotation takes into account data from comparative genomics, such as protein family profiles.

After determining the constraints of the annotation, the YAGA software can automatically annotate *de novo* all genomes from their raw sequences. The predictors used by the YAGA software can also take into account the data RNAseq to reinforce the prediction of genes. The current settings of the software are intended for annotation of the genomes of yeast, but the software is adaptable for all types of species.

5.4. BioRica: Multi-scale Stochastic Modeling

Participants: David James Sherman [correspondant], Rodrigo Assar Cuevas, Alice Garcia.

BioRica is a high-level modeling framework integrating discrete and continuous multi-scale dynamics within the same semantics field. A model in BioRica node is hierarchically composed of nodes, which may be existing models. Individual nodes can be of two types:

- Discrete nodes are composed of states, and transitions described by constrained events, which can be non deterministic. This captures a range of existing discrete formalisms (Petri nets, finite automata, etc.). Stochastic behavior can be added by associating the likelihood that an event fires when activated. Markov chains or Markov decision processes can be concisely described. Timed behavior is added by defining the delay between an event's activation and the moment that its transition occurs.
- Continuous nodes are described by ODE systems, potentially a hybrid system whose internal state flows continuously while having discrete jumps.

The system has been implemented as a distributable software package

The BioRica compiler reads a specification for hierarchical model and compiles it into an executable simulator. The modeling language is a stochastic extension to the AltaRica Dataflow language, inspired by work of Antoine Rauzy. Input parsers for SBML 2 version 4 are curently being validated. The compiled code uses the Python runtime environment and can be run stand-alone on most systems [36].

For more information see biorica.gforge.inria.fr, the BioRica Gforge web site. BioRica was developed as an Inria Technology Development Action (ADT).

5.5. Pathtastic: Inference of whole-genome metabolic models

Participants: David James Sherman [correspondant], Pascal Durrens, Nicolás Loira, Anna Zhukova.

Pathtastic is a software tool for inferring whole-genome metabolic models for eukaryote cell factories. It is based on *metabolic scaffolds*, abstract descriptions of reactions and pathways on which inferred reactions are hung are are eventually connected by an interative mapping and specialization process. Scaffold fragments can be repeatedly used to build specialized subnetworks of the complete model.

Pathtastic uses a consensus procedure to infer reactions from complementary genome comparisons, and an algebra for assisted manual editing of pathways.

For more information see pathtastic.gforge.inria.fr, the Pathtastic Gforge web site.

5.6. Génolevures On Line: Comparative Genomics of Yeasts

Participants: David James Sherman, Pascal Durrens [correspondant], Natalia Golenetskaya, Tiphaine Martin.

The Génolevures online database provides tools and data for exploring the annotated genome sequences of more than 20 genomes, determined and manually annotated by the Génolevures Consortium to facilitate comparative genomic studies of hemiascomycetous yeasts. Data are presented with a focus on relations between genes and genomes: conservation of genes and gene families, speciation, chromosomal reorganization and synteny. The Génolevures site includes an area for specific studies by members of its international community.

Génolevures online uses the MAGUS system for genome navigation, with project-specific extensions developed by David Sherman, Pascal Durrens, and Tiphaine Martin. An advanced query system for data mining in Génolevures is being developed by Natalia Golenetskaya. The contents of the knowledge base are expanded and maintained by the CNRS through GDR 2354 Génolevures. Technical support for Génolevures On Line is provided the CNRS through UMR 5800 LaBRI.

For more information see genolevures.org, the Génolevures web site.

6. New Results

6.1. Yeast comparative genomics

Participants: David James Sherman, Pascal Durrens [correspondant], Tiphaine Martin, Nicolás Loira.

By using the MAGNOME software developments, including the MAGUS system and YAGA software, we have successfully realized a full annotation and analysis of seven new genomes, provided to the Génolevures Consortium by the CEA–Génoscope (Évry). Two distant genomes from the *Debaryomycetaceae* and *mitosporic Saccharomycetales* clades of the *Saccharomycetales* were annotated using previously published Génolevures genomes [5], [9], [10] as references. A further group of five species, comprised of pathogenic and nonpathogenic species, was analyzed with the goal of identifying virulence determinants[13]. By choosing species that are highly related but which differ in the particular traits that are targeted, in this case pathogenicity, we are able to focus of the few hundred genes related to the trait. The approximately 40,000 new genes from these studies were classified into existing Génolevures families as well as branch-specific families. The results from these two studies will be published in the coming year.

6.2. Assembly, annotation and comparison of Oenococcus strains

Participants: David James Sherman, Pascal Durrens, Elisabeth Bon [correspondant], Tiphaine Martin, Aurélie Goulielmakis.

Oenococcus oeni is part of the natural microflora of wine and related environments, and is the main agent of the malolactic fermentation (MLF), a step of wine making that generally follows alcoholic fermentation (AF) and contributes to wine deacidification, improvement of sensorial properties and microbial stability. The start, duration and achievement of MLF are unpredictable since they depend both on the wine characteristics and on the properties of the *O. oeni* strains. In collaboration with Patrick Lucas's lab of the ISVV Bordeaux that is currently proceeding with genome sequencing, explorative and, and comparative genomics, Elisabeth Bon coordinates our efforts into the OENIKITA project (since 2009), a scale switching challenge including highthrouput exploratory and comparative genomics for oenological bacterial starters, and the development of an online web-collaborative multigenomic comparative platform (under development) based on the the Génolevures database architecture and MAGUS / YAGA systems.

OENI-Genomics axis: In comparative genomics, we investigated gene repertoire and genomic organization conservation through intra- and inter-species genomic comparisons, which clearly show that the *O. oeni* genome is highly plastic and fast-evolving. Results reveal that the optimal adaptation to wine of a strain mostly depends on the presence of key adaptive loops and polymorphic genes. They also point up the role of horizontal gene transfer and mobile genetic elements in *O. oeni* genome plasticity, and give the first clues of the genetic origin of its oenological aptitudes. As a result of the scaling out challenge, we completed the assembly of 19 fully sequenced *O. oeni* genome variants.

KITA-Genomics (E. Bon, D. Sherman): This project that is focused on the sequencing, assembly, exploration and comparison of the O. kitaharae genome, has benefited to an international collaboration involving Dr V. Makeev. MAGNOME contributed to the assembly of the genome. The comparison against the O. oeni genomic architecture will contribute to shed light on the evolutionary mechanisms which are responsible for the atypically long branch of the genus Oenococcus in phylogenetic trees.

Transcriptomic axis (E. Bon, A. Goulielmakis): Under the supervision of E. Bon, Aurélie Goulielmakis has completed for the ANR DIVOENI a detailed manual annotation of a new reference strain of *O. oeni* and performed comparative transcriptome analysis to identify genes differentially expressed under different culture conditions. We explored and compared how the expression system is solicited when *O. oeni* strains adapted to grow in some niches are placed under stress-exposure conditions. The monitoring of gene expression status between strains, through the definition of a global expression pattern proper to each gene, partially lift the veil on how *O. oeni* genome adapts function to its environment. The weight of genetic background and ecological niche pressure on gene expression flexibility was evaluated, and the *O. oeni* pan-transcriptome architecture characterized. The first guidelines revealed a supra-spatial organization of stress response into activated and repressed larger macro-domains defining functional landmarks and intra-chromosomal territories [16]. Decryption of stress-sensitive gene repertoires promises to be an efficient tool in the conquest of *O. oeni* "domestication" through the identification of molecular markers responsible for different physiological capabilities, and the selection of the best adapted strains.

Gene plasticity modelisation (E. Bon, A. Goulielmakis): A novel axis of research recently emerged under the initiative of E. Bon (pseudOE project) around the detection, characterization and conservation of pseudogenes populations in Oenococcus bacteria. Such topic presents a double interest: phylogenetic at first because it should allow to better estimate the degree of genic/genomic plasticity of these bacteria, and algorithmic then because the pseudogenes are a source of confusion for the automatic prediction of genes. Through a transversal collaboration and a cooperative supervision with the Algorithms for Analysis of Biological Structures Group (P. Ferraro, J. Allali) at LaBRI, Laetitia Bourgeade (PhD, Univ. Bordeaux1) was recruited to develop dedicated methods to improve pseudogenes automatic detection, and therefore gene predictions, and to reconstruct fossil and modern genes evolutionary history.

6.3. Scaling-out

Participants: David James Sherman [correspondant], Pascal Durrens, Tiphaine Martin, Natalia Golenetskaya, Florian Lajus.

The Tsvetok project in MAGNOME targets "scaling out" for data and computation, both to improve capacity for handling large volumes of data and to permit more automatic analysis of projects of the "comparative genomics of related species" type, where a set of genomes is sequenced and analyzed as part of the same process. Natalia Golenetskaya has designed and implemented a NoSQL schema through the identification of standard queries, definition of the appropriate query-oriented storage schema, and mapping of structured values to this schema. This prototype is being tested on an Apache Cassandra ring deployed in MAGNOME's dedicated computing cluster.

Large-scale data-mining such as that required for comparative genomics is fundamentally *data-parallel*: an initial transformation is applied to every data object of a given type (such as genes or even individual nucleotides), then a statistical machine learning procedure is applied to the transformed data to produce a summary or to learn a classification function. Analyses of this kind are the design goal of the MapReduce paradigm [31]. Using Tsvetok as a generator for Apache Hadoop, Natalia is designing MapReduce solutions for the principal whole-genome and data-mining analyses used by MAGNOME for eukaryote and prokaryote comparative genomics.

6.4. Affinity Proteomics: Standards for affinity binders

Participants: David James Sherman [correspondant], Natalia Golenetskaya.

Last year we successfully completed and released the MIAPAR and PSI-PAR international standards for knowledge representation and data exchange of affinity binder properties, a five-year effort organized as part of the ProteomeBinders and HUPO-PSI consortia. These standards were reported in *Nature Biotechnology* and *Molecular and Cellular Proteomics* to the research community [3] [37], [52] and extend previous work [38], [43]. One long-standing issue is the adoption of these standards by individual researchers in the lab: initial data entry must be simple enough that standards-based reporting can be integrated into the process of writing the paper. We used an extensive dataset of affinity proteomics data to evaluate "last mile" tools for data entry and initial reporting of affinity proteomics data, and identified places where existing tools need to be adapted to meet these specific needs[21].

6.5. Inferring metabolic models

Participants: David James Sherman [correspondant], Pascal Durrens, Tiphaine Martin, Nicolás Loira, Anna Zhukova.

In collaboration with Prof Jean-Marc Nicaud's lab at the INRA Grignon, we developed the first functional genome-scale metabolic model of an oleaginous yeast. Most work in producing genome-scale metabolic models has focused on model organisms, in part due to the cost of obtaining well-annotated genome sequences and sufficiently complete experimental data for refining and verifying the models. However, for many fungal genomes of biotechnological interest, the combination of large-scale sequencing projects and in-depth experimental studies has made it feasible to undertake metabolic network reconstruction for a wider range of organisms.

An excellent representative of this new class of organisms is *Yarrowia lipolytica*, an oleaginous yeast studied experimentally for its role as a food contaminant and its use in bioremediation and cell factory applications. As one of the hemiascomycetous yeasts completely sequenced in the Génolevures program it enjoys a high quality manual annotation by a network of experts. It is also an ideal subject for studying the role of species-specific expansion of paralogous families, a considerable challenge for eukaryotes in genome-scale metabolic construction. To these ends, we undertook a complete reconstruction of the *Y. lipolytica* metabolic network.

Methods: A draft model was extrapolated from the *S. cerevisiae* model iIN800, using *in silico* methods including enzyme conservation predicted using Génolevures and reaction mapping maintaining compartments. This draft was curated by a group of experts in *Y. lipolytica* metabolism, and iteratively improved and validated through comparison with experimental data by flux balance analysis. Gap filling, species-specific reactions, and the addition of compartments with the corresponding transport reactions were among the improvements that most affected accuracy.

Results: We produced an accurate functional model for *Y. lipolytica*, MODEL1111190000 in Biomodels.net, that has been qualitatively validated against gene knockouts.

6.6. Hierarchical modeling with BioRica

Participants: David James Sherman [correspondant], Tiphaine Martin, Alice Garcia, Rodrigo Assar-Cuevas, Nicolás Loira.

A recurring challenge for *in silico* modeling of cell behavior is that experimentally validated models are so focused in scope that it is difficult to repurpose them. Hierarchical modeling is one way of combining specific models into networks. Effective use of hierarchical models requires both formal definition of the semantics of such composition, and efficient simulation tools for exploring the large space of complex behaviors.

BioRica is a high-level hierarchical modeling framework for models combining continuous and discrete components. By providing a reliable and functional software tool backed by a rigorous semantics, we hope to advance real adoption of hierarchical modeling by the systems biology community. By providing an understandable and mathematically rigorous semantics, this will make is easier for practicing scientists to build practical and functional models of the systems they are studying, and concentrate their efforts on the system rather than on the tool.

Rodrigo Assar formalized two strategies for integrating discrete control with continuous models, coefficient switches that control the parameters of the continuous model, and strong switches that choose different models. This was translated by Alice Garcia into a BioRica specification for hybrid systems that assures integrity of models, allowing composition, reconciliation, and reuse of models with SBML specifications. Rodrigo used this approach to describe two systems: wine fermentation kinetics, and cell fate decisions leading to bone and fat formation[11]. In the first, known models that describe the responses of yeast cells to different temperatures, resources and toxins, were reconciled using coefficient switches that gave the best adjustment of the model depending on the initial conditions and fermentation variable. In the second, a combination of accurate models to predict the bone and fat formation in response to activation of pathways such as the Wnt pathway, and changes of conditions affecting these functions such as increments in Homocysteine, were used to analyze the responses to treatments for osteoporosis and other bone mass disorders. Our hope is that this is a first step in obtaining *in silico* evaluations of medical treatments before testing them *in vivo* or *in vitro*.

Maria Llubères of the University of Puerto Rico visited MAGNOME and we established formal relationships between BioRica models and probabilistic boolean networks.

7. Contracts and Grants with Industry

7.1. Contracts with Industry

SARCO, the research subsidiary of the Laffort group, has entered into a contract with MAGNOME to develop comparative genomics tools for selecting wine starters. This contract will permit SARCO to take a decisive step in the understanding of oenological microorganisms by obtaining and exploiting the sequences of their genomes. Comparison of the genomes of these strains has become absolutely necessary for learning the genetic origin of the phenotypic variations of oenological yeasts and bacteria. This knowledge will permit SARCO to optimize and accelerate the process of selection of the highest-performing natural strains. With the help of MAGNOME members and their rich experience in comparative analysis of related genomes, SARCO will acquire competence in biological analysis of genomic sequences. At the same time, MAGNOME members will acquire further experience with the genomes of winemaking microorganisms, which will help us define new tools and methods better adapted to this kind of industrial cell factory.

7.2. Grants with Industry

The French Petroleum Institute (*Institut français de pétrole-énergies nouvelles*) is coordinating a 6 M-Euro contract with the Civil Aviation Directorate (*Direction Générale de l'Aviation Civile*) on behalf of a large consortium of industrial (EADS, Dassault, Snecma, Turbomeca, Airbus, Air France, Total) and academic (CNRS, INRA, Inria) partners to explore different technologies for alternative fuels for aviation. The CAER project studies both biofuel products and production, improved jet engine design, and the impact of aircraft. Within CAER MAGNOME via CNRS, works with partners from Grignon and Toulouse on the genomics of highly-performant oleaginous yeasts.

8. Partnerships and Cooperations

8.1. Regional Initiatives

8.1.1. Aquitaine Region "SAGÉSS" comparative genomics for wine starters

Participants: David James Sherman [correspondant], Pascal Durrens, Elisabeth Bon, Tiphaine Martin, Nicolás Loira.

This project is a collaboration between the company SARCO, specialized in the selection of industrial yeasts with distinct technological abilities, with the ISVV and MAGNOME. The goal is to use genome analysis to identify molecular markers responsible for different physiological capabilities, as a tool for selecting yeasts and bacteria for wine fermentation through efficient hybridization and selection strategies. This collaboration has obtained the INNOVIN label.

8.1.2. Aquitaine Region "Oenophages: comparative genomics for oenococcus bacteriophages" (2011-2014)

Participants: David James Sherman [correspondant], Elisabeth Bon.

8.2. National Initiatives

8.2.1. ANR DIVOENI, 2008-2012

Participants: Elisabeth Bon [correspondant], Aurélie Goulielmakis.

Elisabeth Bon is a partner in DIVOENI, a four-year ANR project concerning intraspecies biodiversity of the oenological bacteria *Oenococcus oeni*. Coordinated by Prof. Aline Lonvaud (Univ. Bordeaux Segalen) from the Institute of Vine and Wine Sciences of Bordeaux – Aquitaine, this scientific programme was developed:

- 1. To evaluate the genetic diversity of a vast collection of strains, to set up phylogenetic groups, then to investigate relationships between the ecological niches (cider, wine, champagne) and the essential phenotypical traits. Hypotheses on the evolution in the species and on the genetic stability of strains will be drawn.
- 2. To propose methods based on molecular markers to make a better use of the diversity of the species.
- 3. To measure the impact of the repeated use of selected strains on the diversity in the ecosystem and to draw the conclusions for its preservation.

Elisabeth is in charge of the computational infrastructure dedicated to genomics and post-genomics data storage, handling and analysis. She coordinates collaboration with the CBiB-Centre de Bioinformatique de Bordeaux.

8.3. European Initiatives

8.3.1. Affinity Proteomics (FP7)

Participants: David James Sherman [correspondant], Natalia Golenetskaya.

A major objective of the "post-genome" era is to detect, quantify and characterise all relevant human proteins in tissues and fluids in health and disease. This effort requires a comprehensive, characterised and standardised collection of specific ligand binding reagents, including antibodies, the most widely used such reagents, as well as novel protein scaffolds and nucleic acid aptamers. Currently there is no pan-European platform to coordinate systematic development, resource management and quality control for these important reagents.

MAGNOME is an associate partner of the FP7 "Affinity Proteome" project coordinated by Prof. Mike Taussig of the Babraham Institute and Cambridge University. Within the consortium, we participate in defining community for data representation and exchange, and evaluate knowledge engineering tools for affinity proteomics data.

8.3.2. Sustained Collaborations with Major European Organization

Prof. Mike Taussig: Babraham Institute & Cambridge University

Knowledge engineering for Affinity Proteomics

Henning Hermjakob: European Bioinformatics Institute

Standards and databases for molecular interactions

8.4. International Initiatives

8.4.1. Visits of International Scientists

Participants: Vsevolod Makeev, Artëm Kasianov, Marie Llubères.

Vsevolod Makeev (Senior Researcher at the Russian Academy of Sciences, Vavilov Institute) has been a collaborator for several years. He and his student Artëm Kasianov made several visits to Inria in 2011, and worked with us on genome assembly algorithms, computational identification of sequence motifs, and distributed algorithms for data mining. Vsevolod Makeev was a visiting CNRS Senior Researcher in the LaBRI and MAGNOME for three months in the Fall of 2011.

Marie Llubères visited MAGNOME from the University of Puerto Rico for two months on a grant from the NSF PIRE program. She worked on hierarchical modeling of biological systems and specifically on bijections between Probabilistic Boolean Networks and the Stochastic Transition Systems used in the BioRica framework.

8.4.1.1. Internship

Participant: Hugo Campbell Sills.

Hugo Campbell Sills came to MAGNOME on an Inria International Internship in the Summer of 2011, and worked on single-nucleotide polymorphism discovery and effects in twelve œnological yeast genomes.

8.4.2. Participation In International Programs

8.4.2.1. Génolevures and Dikaryome Consortia

Participants: David James Sherman [correspondant], Pascal Durrens, Tiphaine Martin, Nicolás Loira, Anasua Sarkar, Anna Zhukova, Florian Lajus.

Since 2000 our team is a member of the Génolevures Consortium (GDR CNRS), a large-scale comparative genomics project that aims to address fundamental questions of molecular evolution through the sequencing and the comparison of 14 species of hemiascomycetous yeasts. The Consortium is comprised of 16 partners, in France, Belgium, Spain, the Netherlands (see http://genolevures.org/). Within the Consortium, our team is responsible for bioinformatics, for research in new methods of analysis. Pascal Durrens and Tiphaine Martin of the CNRS are responsible for the development of resources for exploiting comparative genomic data. Pascal Durrens is the editorial manager of the Génolevures on-line resource.

The Dikaryome Consortium is a scientific collaboration between several international partners and the National Center for Sequencing (CEA–Génoscope, Évry) on the sequencing, annotation, and comparative analysis of fungal genomes.

These perennial collaborations continue in two ways. First, a number of new projects are underway, concerning several new genomes currently being sequenced, and new questions about the mechanisms of gene formation. Second, through the development and improvement of the Génolevures On Line database, in whose maintenance our team has a longstanding committment.

9. Dissemination

9.1. Animation of the scientific community

David Sherman is member of the editorial board of the journal *Computational and Mathematical Methods in Medicine*, and reviewer for several in the bioinformatics field.

David Sherman was external reviewer and member of the thesis defense jury for Anne-Ruxandra Carvunis, Grenoble. He was a member and president of the jury for the thesis defense of Anne-Laure Gaillard, Bordeaux. He was thesis director and member of the jury for the thesis defense of Rodrigo Assar. He was a member of the HDR defense jury for Patrick Lucas, Bordeaux.

Pascal Durrens is responsible for scientific diffusion, and David Sherman is head of Bioinformatics, for the Génolevures Consortium.

Pascal Durrens is leader of the "Comparative Genomics" theme and member of the Scientific Council of the LaBRI UMR 5800/CNRS

Tiphaine Martin is member of the Local Committee and member of Local Committee for Occupational Health and Safety of the INRIA Bordeaux Sud-Ouest.

Tiphaine Martin is member of the GIS-IBiSA GRISBI-Bioinformatics Grid working group.

Tiphaine Martin and David Sherman are members of the *Institut de Grilles*, and Tiphaine is active in the Biology/Health working group.

Elisabeth Bon is member of the "Comité Technique Paritaire" (2008-2011) and the "Comité Technique de Proximité" (since 2011-10-20) at the Segalen Bordeaux University

9.2. Teaching

David Sherman and Natalia Golenetskaya teach :

Master : Web et Interfaces Homme-Machine, 50h, 2ème année Ingénieur, Enseirb-Matmeca (Institut Polytechnique de Bordeaux), Bordeaux

Tiphaine Martin teaches :

Master : Utilisation of EGEE GRID via virtual organisation GRISBI , 8h, niveau (M2), University Lyon, France

Master : Utilisation of EGEE GRID via virtual organisation GRISBI, 8h, niveau (M2), INRA Toulouse, France

Doctorat : Utilisation of MAGUS software, 8h, Génolevures Consortium, France

Tiphaine Martin has the supervision of 4 Bioinformatics MSc students from the University of Bordeaux:

Master : Development of search tools on Ge´nolevures databases, 6hETC, M1, University Bordeaux 1 and University Bordeaux Segalen, France

Elisabeth Bon is Associate Professor in Bioinformatics and Genomics, and teaches undergraduate courses in computer sciences in regular STS (Sciences, Technologies & Sante) bachelor's degrees and research oriented STS master's degrees at the Life Sciences Department of the University Bordeaux Segalen (Medicine and Life Sciences schools) and University Bordeaux 1 (Computer and Life Science schools).

Licence : "Introduction to ICTs-Information & Communication Technologies" class (basic and advanced sections), 112H00, niveau (L1, L2), the STS- biology variant Licence program, université, France

Licence : the national "IT and Internet certificate (C2i®), level 1", 20h, niveau (L2, L3), the STSbiology variant Licence program, université, France

Master : "Bioinformatics: Computerised resources, data banks and methods", 60h, niveau (M1),the Biology & Healthcare STS- Master program, co-listed with the University Bordeaux 1 (Sciences & Technologies) and the University Bordeaux Segalen (Medicine & Life Sciences), France

Elisabeth Bon is :

Licence : Responsible for the bachelor's degree "Information Technologies & Internet advanced course", Life Sciences Department, University Bordeaux, France

Licence : Responsible for the "IT and Internet certificate (C2i®), level 1", Life Sciences Department, University Bordeaux , France

Licence : Current president (2005-2007; since sept. 2011) of the "Segalen Bordeaux University IT and Internet certificate (C2i, level 1) committee" in charge of the C2i evaluation and certification for students and continuous education interns, University Bordeaux Segalen, France

Master : Master Theses in Computer Science, speciality in BioInformatics: L. Bourgeade (2011-02-01 / 2011-08-31), Reconstitution in silico de l'histoire évolutive des pseudogènes chez les bactéries lactiques du genre Oenococcus, M2, University Bordeaux 1, France

Doctorat : Ph.D. Thesis in Computer Science: L. BOURGEADE (Since 2011-10-01) in cooperation with P. Ferraro and J. Allali at LaBRI, Filtres sur les arborescences modélisant les ARN et plasticité génique, University Bordeaux 1, France

Doctorat : ATER (assistant professor) in computer sciences for ITCs practical workshops and courses in the first year of the Bachelor's degre, University Bordeaux 1, France

PhD & HdR:

PhD: Rodrigo Assar, Modeling and simulation of Hybrid Systems and Cell factory applications, University Bordeaux 1, defended 2011-10-26, David Sherman

PhD in progress: Nicolás Loira, University Bordeaux 1, Scaffold-based Reconstruction Method for Genome-Scale Metabolic Models, began 2007, David Sherman

PhD in progress: Natalia Golenetskaya, University Bordeaux 1, began 2009, Scaling out for data in comparative genomics, Pascal Durrens and David Sherman

PhD in progress : Razanne Issa, University Bordeaux 1, Analyse symbolique de données génomiques, began 2010, David Sherman

PhD in progress: Anna Zhukova, University Bordeaux 1, Comparative genomics of biotechnological organisms, began 2011, David Sherman

PhD in progress: Anasua Sarkar, University Bordeaux 1, began 2008, Macha Nikolski

PhD in progress: Laetitia Bourgeade, University Bordeaux 1, Filtres sur les arborescences modélisant les ARN et plasticité génique, began, Pascal Ferarro and Elisabeth Bon

10. Bibliography

Major publications by the team in recent years

- [1] R. BARRIOT, D. J. SHERMAN, I. DUTOUR. *How to decide which are the most pertinent overly-represented features during gene set enrichment analysis*, in "BMC Bioinformatics", 2007, vol. 8 [DOI: 10.1186/1471-2105-8-332], http://hal.inria.fr/inria-00202721/en/.
- [2] G. BLANDIN, P. DURRENS, F. TEKAIA, M. AIGLE, M. BOLOTIN-FUKUHARA, E. BON, S. CASAREGOLA, J. DE MONTIGNY, C. GAILLARDIN, A. LÉPINGLE, B. LLORENTE, A. MALPERTUY, C. NEUVÉGLISE, O. OZIER-KALOGEROPOULOS, A. PERRIN, S. POTIER, J.-L. SOUCIET, E. TALLA, C. TOFFANO-NIOCHE, M. WÉSOLOWSKI-LOUVEL, C. MARCK, B. DUJON. Genomic Exploration of the Hemiascomycetous Yeasts: 4. The genome of Saccharomyces cerevisiae revisited, in "FEBS Letters", December 2000, vol. 487, n^o 1, p. 31-36.
- [3] J. BOURBEILLON, S. ORCHARD, I. BENHAR, C. BORREBAECK, A. DE DARUVAR, S. DÜBEL, R. FRANK, F. GIBSON, D. GLORIAM, N. HASLAM, T. HILTKER, I. HUMPHREY-SMITH, M. HUST, D. JUNCKER, M. KOEGL, Z. KONTHUR, B. KORN, S. KROBITSCH, S. MUYLDERMANS, P.-A. NYGREN, S. PALCY, B. POLIC, H. RODRIGUEZ, A. SAWYER, M. SCHLAPSHY, M. SNYDER, O. STOEVESANDT, M. J. TAUSSIG, M. TEMPLIN, M. UHLEN, S. VAN DER MAAREL, C. WINGREN, H. HERMJAKOB, D. J. SHERMAN. *Minimum information about a protein affinity reagent (MIAPAR).*, in "Nature Biotechnology", 07 2010, vol. 28, n⁰ 7, p. 650-3 [DOI: 10.1038/NBT0710-650], http://hal.inria.fr/inria-00544750/en.
- [4] A. B. CANELAS, N. HARRISON, A. FAZIO, J. ZHANG, J.-P. PITKÄNEN, J. VAN DEN BRINK, B. M. BAKKER, L. BOGNER, J. BOUWMAN, J. I. CASTRILLO, A. CANKORUR, P. CHUMNANPUEN, P. DARAN-LAPUJADE, D. DIKICIOGLU, K. VAN EUNEN, J. C. EWALD, J. J. HEIJNEN, B. KIRDAR, I. MATTILA, F. I. C. MENSONIDES, A. NIEBEL, M. PENTTILÄ, J. T. PRONK, M. REUSS, L. SALUSJÄRVI, U. SAUER, D. J. SHERMAN, M. SIEMANN-HERZBERG, H. WESTERHOFF, J. DE WINDE, D. PETRANOVIC, S. G. OLIVER, C. T. WORKMAN, N. ZAMBONI, J. NIELSEN. Integrated multilaboratory systems biology reveals differences in protein metabolism between two reference yeast strains., in "Nature Communications", 12 2010, vol. 1, n^O 9, 145 [DOI: 10.1038/NCOMMS1150], http://hal.inria.fr/inria-00562005/en/.
- [5] B. DUJON, D. J. SHERMAN, G. FISCHER, P. DURRENS, S. CASAREGOLA, I. LAFONTAINE, J. DE MONTIGNY, C. MARCK, C. NEUVÉGLISE, E. TALLA, N. GOFFARD, L. FRANGEUL, M. AIGLE, V. ANTHOUARD, A. BABOUR, V. BARBE, S. BARNAY, S. BLANCHIN, J.-M. BECKERICH, E. BEYNE, C. BLEYKASTEN, A. BOISRAMÉ, J. BOYER, L. CATTOLICO, F. CONFANIOLERI, A. DE DARUVAR, L. DESPONS, E. FABRE, C. FAIRHEAD, H. FERRY-DUMAZET, A. GROPPI, F. HANTRAYE, C. HENNEQUIN, N. JAUNIAUX, P. JOYET, R. KACHOURI-LAFOND, A. KERREST, R. KOSZUL, M. LEMAIRE, I. LESUR, L. MA, H. MULLER, J.-M. NICAUD, M. NIKOLSKI, S. OZTAS, O. OZIER-KALOGEROPOULOS, S. PELLENZ, S. POTIER, G.-F. RICHARD, M.-L. STRAUB, A. SULEAU, D. SWENNEN, F. TEKAIA, M. WÉSOLOWSKI-LOUVEL, E. WESTHOF, B. WIRTH, M. ZENIOU-MEYER, I. ZIVANOVIC, M. BOLOTIN-FUKUHARA, A. THIERRY, C. BOUCHIER, B. CAUDRON, C. SCARPELLI, C. GAILLARDIN, J. WEISSENBACH, P. WINCKER, J.-L. SOUCIET. Genome evolution in yeasts, in "Nature", 07 2004, vol. 430, n^O 6995, p. 35-44 [DOI: 10.1038/NATURE02579], http://hal.archives-ouvertes.fr/hal-00104411/en/.
- [6] P. DURRENS, M. NIKOLSKI, D. J. SHERMAN. Fusion and fission of genes define a metric between fungal genomes., in "PLoS Computational Biology", 10 2008, vol. 4, e1000200 [DOI: 10.1371/JOURNAL.PCBI.1000200], http://hal.inria.fr/inria-00341569/en/.

- [7] A. GOËFFON, M. NIKOLSKI, D. J. SHERMAN. An Efficient Probabilistic Population-Based Descent for the Median Genome Problem, in "Proceedings of the 10th annual ACM SIGEVO conference on Genetic and evolutionary computation (GECCO 2008)", Atlanta United States, ACM, 2008, p. 315-322, http://hal. archives-ouvertes.fr/hal-00341672/en/.
- [8] M. NIKOLSKI, D. J. SHERMAN. Family relationships: should consensus reign?- consensus clustering for protein families, in "Bioinformatics", 2007, vol. 23, p. e71–e76 [DOI: 10.1093/BIOINFORMATICS/BTL314], http://hal.inria.fr/inria-00202434/en/.
- [9] D. J. SHERMAN, T. MARTIN, M. NIKOLSKI, C. CAYLA, J.-L. SOUCIET, P. DURRENS. Genolevures: protein families and synteny among complete hemiascomycetous yeast proteomes and genomes., in "Nucleic Acids Research (NAR)", 2009, p. D550-D554 [DOI: 10.1093/NAR/GKN859], http://hal.inria.fr/inria-00341578/ en/.
- [10] J.-L. SOUCIET, B. DUJON, C. GAILLARDIN, M. JOHNSTON, P. V. BARET, P. CLIFTEN, D. J. SHER-MAN, J. WEISSENBACH, E. WESTHOF, P. WINCKER, C. JUBIN, J. POULAIN, V. BARBE, B. SÉGURENS, F. ARTIGUENAVE, V. ANTHOUARD, B. VACHERIE, M.-E. VAL, R. S. FULTON, P. MINX, R. WILSON, P. DURRENS, G. JEAN, C. MARCK, T. MARTIN, M. NIKOLSKI, T. ROLLAND, M.-L. SERET, S. CASARE-GOLA, L. DESPONS, C. FAIRHEAD, G. FISCHER, I. LAFONTAINE, V. LEH, M. LEMAIRE, J. DE MON-TIGNY, C. NEUVÉGLISE, A. THIERRY, I. BLANC-LENFLE, C. BLEYKASTEN, J. DIFFELS, E. FRITSCH, L. FRANGEUL, A. GOËFFON, N. JAUNIAUX, R. KACHOURI-LAFOND, C. PAYEN, S. POTIER, L. PRIBYLOVA, C. OZANNE, G.-F. RICHARD, C. SACERDOT, M.-L. STRAUB, E. TALLA. Comparative genomics of protoploid Saccharomycetaceae., in "Genome Research", 2009, vol. 19, p. 1696-1709, http://hal.inria.fr/inria-00407511/en/.

Publications of the year

Doctoral Dissertations and Habilitation Theses

[11] R. ASSAR. *Modeling and simulation of Hybrid Systems and Cell factory applications*, Université Sciences et Technologies - Bordeaux I, October 2011, http://hal.inria.fr/tel-00635273/en.

Articles in International Peer-Reviewed Journal

- [12] P. DURRENS, T. MARTIN, D. J. SHERMAN. *The Génolevures database*, in "Comptes Rendus de l'Académie des Sciences, Série Biologies", 2011, http://hal.inria.fr/inria-00539200/en.
- [13] A. ENACHE-ANGOULVANT, J. GUITARD, F. GRENOUILLET, T. MARTIN, P. DURRENS, C. FAIRHEAD, C. HENNEQUIN. Rapid Discrimination between Candida glabrata, Candida nivariensis, and Candida bracarensis by Use of a Singleplex PCR, in "Journal of Clinical Microbiology", September 2011, vol. 49, n^o 9, p. 3375-3379 [DOI: 10.1128/JCM.00688-11], http://hal.inria.fr/inria-00625115/en.

Invited Conferences

- [14] T. MARTIN. *Genolevures: automated annotation of yeast genome sequences*, in "Comparative Genomics of Eukaryotic Microorganisms", Sant Feliu de Guixols, Spain, October 2011, http://hal.inria.fr/hal-00640571/en.
- [15] D. J. SHERMAN, N. GOLENETSKAYA. Addressing scaling-out challenges for comparative genomics, in "Moscow Conference on Computational Molecular Biology", Moscow, Russian Federation, July 2011, http:// hal.inria.fr/hal-00649189/en.

International Conferences with Proceedings

- [16] A. GOULIELMAKIS, J. BRIDIER, A. BARRÉ, O. CLAISSE, D. J. SHERMAN, P. DURRENS, A. LONVAUD-FUNEL, E. BON. How does Oenococcus oeni adapt to its environment? A pangenomic oligonucleotide microarray for analysis O. oeni gene expression under wine shock., in "OENO2011- 9th International Symposium of Oenology", Bordeaux, France, Dunod, Paris, January 2011, http://hal.inria.fr/hal-00646867/en.
- [17] T. MARTIN, P. DURRENS. *Génolevures: Policy for Automated Annotation of Genome Sequences*, in "JOBIM 2011", Paris, France, June 2011, http://hal.inria.fr/inria-00614485/en.
- [18] T. MARTIN, D. J. SHERMAN, P. DURRENS. Genolevures : automated annotation of yeast genome sequences, in "Comparative Genomics of Eukaryotic Microorganisms", Sant Feliu de Guixols, Spain, October 2011, http://hal.inria.fr/hal-00640575/en.

National Conferences with Proceeding

- [19] R. ASSAR, A. GARCIA, D. J. SHERMAN. Modeling Stochastic Switched Systems with BioRica, in "Journées Ouvertes en Biologie, Informatique et Mathématiques JOBIM 2011", Paris, France, Institut Pasteur, July 2011, p. 297–304, http://hal.inria.fr/inria-00617419/en.
- [20] C. BLANCHET, C. GAUTHEY, C. CARON, O. COLLIN, S. DELMOTTE, T. MARTIN, A. ROULT, F. SAMSON, B. SPATARO. *RENABI GRISBI Infrastructure Distribuée pour la Bioinformatique*, in "JOBIM 2011 - Journées Ouvertes Biologie Informatique Mathématique", Paris, France, June 2011, http://hal.inria.fr/hal-00640007/en.

Conferences without Proceedings

- [21] N. GOLENETSKAYA, D. J. SHERMAN. Assessing "last mile" tools for affinity binder databases, in "5th ESF Workshop on Affinity Proteomics: Ligand Binders against the Human Proteome", Alpbach, Austria, March 2011, http://hal.inria.fr/hal-00653518/en.
- [22] N. GOLENETSKAYA, D. J. SHERMAN. Rethinking global analyses and algorithms for comparative genomics in a functional MapReduce style, in "Algorithmique, combinatoire du texte et applications en bio-informatique (SeqBio 2011)", Lille, France, December 2011, http://hal.inria.fr/hal-00654797/en.
- [23] T. MARTIN, P. DURRENS. Un polymorphisme suspect, in "Unithé ou Café", Talence, France, June 2011, http://hal.inria.fr/inria-00614484/en.
- [24] D. J. SHERMAN, N. GOLENETSKAYA, T. MARTIN, P. DURRENS. Comparative annotation and scalingout challenges for paraphyletic strategies, in "EMBO Symposium on Comparative Genomics of Eukaryotic Microorganisms: Understanding the Complexity of Diversity", San Feliu de Guixols, Spain, EMBO, October 2011, http://hal.inria.fr/hal-00652903/en.

References in notes

[25] R. ASSAR, F. VARGAS, D. J. SHERMAN. Reconciling competing models: a case study of wine fermentation kinetics, in "Algebraic and Numeric Biology 2010", Austria Hagenberg, K. HORIMOTO, M. NAKATSUI, N. POPOV (editors), Research Institute for Symbolic Computation, Johannes Kepler University of Linz, 08 2010, p. 68–83, http://hal.inria.fr/inria-00541215/en.

- [26] A. ATHANE, E. BILHÈRE, E. BON, P. LUCAS, G. MOREL, A. LONVAUD-FUNEL, C. LE HÉNAFF-LE MARREC. Characterization of an acquired-dps-containing gene island in the lactic acid bacterium Oenococcus oeni, in "Journal of Applied Microbiology", 2008, Received 22 October 2007, revised 8 April 2008 & Accepted 8 May 2008 (In press), http://hal.inria.fr/inria-00340058/en/.
- [27] R. BARRIOT, J. POIX, A. GROPPI, A. BARRE, N. GOFFARD, D. J. SHERMAN, I. DUTOUR, A. DE DARUVAR. New strategy for the representation and the integration of biomolecular knowledge at a cellular scale, in "Nucleic Acids Research (NAR)", 2004, vol. 32, p. 3581-9 [DOI: 10.1093/NAR/GKH681], http:// hal.inria.fr/inria-00202722/en/.
- [28] E. BON, C. GRANVALET, F. REMIZE, D. DIMOVA, P. LUCAS, D. JACOB, A. GROPPI, S. PENAUD, C. AULARD, A. DE DARUVAR, A. LONVAUD-FUNEL, J. GUZZO. *Insights into genome plasticity of the wine-making bacterium Oenococcus oeni strain ATCC BAA-1163 by decryption of its whole genome.*, in "9th Symposium on Lactic Acid Bacteria", Egmond aan Zee Netherlands, 2008, http://hal.inria.fr/inria-00340073/en/.
- [29] P. CLIFTEN, P. SUDARSANAM, A. DESIKAN, L. FULTON, R. S. FULTON, J. MAJORS, R. WATERSTON, B. A. COHEN, M. JOHNSTON. *Finding functional features in Saccharomyces genomes by phylogenetic footprinting*, in "Science", 2003, vol. 301, p. 71–76.
- [30] M. CVIJOVIC, H. SOUEIDAN, D. J. SHERMAN, E. KLIPP, M. NIKOLSKI. Exploratory Simulation of Cell Ageing Using Hierarchical Models, in "19th International Conference on Genome Informatics Genome Informatics", Gold Coast, Queensland Australia, J. ARTHUR, S.-K. NG (editors), Genome Informatics, Imperial College Press, London, 2008, vol. 21, p. 114–125, EU FP6 Yeast Systems Biology Network LSHG-CT-2005-018942, EU Marie Curie Early Stage Training (EST) Network "Systems Biology", ANR-05-BLAN-0331-03 (GENARISE), http://hal.inria.fr/inria-00350616.
- [31] J. DEAN, S. GHEMAWAT. MapReduce: simplified data processing on large clusters, in "Proceedings of the 6th conference on Symposium on Operating Systems Design and Implementation (OSDI'04)", San Francisco, CA, 2004.
- [32] F. S. DIETRICH, ET AL. . The Ashbya gossypii genome as a tool for mapping the ancient Saccharomyces cerevisiae genome, in "Science", 2004, vol. 304, p. 304-7.
- [33] D. DIMOVA, E. BON, P. LUCAS, R. BEUGNOT, M. DE LEEUW, A. LONVAUD-FUNEL. The whole genome of Oenococcus strain IOEB 8413, in "9th Symposium on Lactic Acid Bacteria", Egmond aan Zee Netherlands, 2008, http://hal.inria.fr/inria-00340086/en/.
- [34] P. FICKERS, A. MARTY, J.-M. NICAUD. The lipases from Yarrowia lipolytica: genetics, production, regulation, biochemical characterization and biotechnological applications, in "Biotechnol Adv.", Nov-Dec 2011, vol. 29, n^o 6, p. 632–44.
- [35] R. FIELDING, R. TAYLOR. Principled design of the modern Web architecture, in "ACM Trans. Internet Technol.", 2002, vol. 2, p. 115–150.
- [36] A. GARCIA, D. J. SHERMAN. *Mixed-formalism hierarchical modeling and simulation with BioRica*, in "11th International Conference on Systems Biology (ICSB 2010)", United Kingdom Edimbourg, 10 2010, P02.446, Poster, http://hal.inria.fr/inria-00529669/en.

- [37] D. GLORIAM, S. ORCHARD, D. BERTINETTI, E. BJÖRLING, E. BONGCAM-RUDLOFF, C. BORREBAECK, J. BOURBEILLON, A. R. M. BRADBURY, A. DE DARUVAR, S. DÜBEL, R. FRANK, T. J. GIBSON, L. GOLD, N. HASLAM, F. W. HERBERG, T. HILTKER, J. D. HOHEISEL, S. KERRIEN, M. KOEGL, Z. KONTHUR, B. KORN, U. LANDEGREN, L. MONTECCHI-PALAZZI, S. PALCY, H. RODRIGUEZ, S. SCHWEINSBERG, V. SIEVERT, O. STOEVESANDT, M. J. TAUSSIG, M. UEFFING, M. UHLÉN, S. VAN DER MAAREL, C. WINGREN, P. WOOLLARD, D. J. SHERMAN, H. HERMJAKOB. A community standard format for the representation of protein affinity reagents., in "Mol Cell Proteomics", 01 2010, vol. 9, n^o 1, p. 1-10 [DOI: 10.1074/MCP.M900185-MCP200], http://hal.inria.fr/inria-00544751/en.
- [38] H. HERMJAKOB, L. MONTECCHI-PALAZZI, G. BADER, J. WOJCIK, L. SALWINSKI, A. CEOL, S. MOORE, S. ORCHARD, U. SARKANS, C. VON MERING, B. ROECHERT, S. POUX, E. JUNG, H. MERSCH, P. KERSEY, M. LAPPE, Y. LI, R. ZENG, D. RANA, M. NIKOLSKI, H. HUSI, C. BRUN, K. SHANKER, S. GRANT, C. SANDER, P. BORK, W. ZHU, A. PANDEY, A. BRAZMA, B. JACQ, M. VIDAL, D. J. SHERMAN, P. LEGRAIN, G. CESARENI, I. XENARIOS, D. EISENBERG, B. STEIPE, C. HOGUE, R. APWEILER. *The HUPO PSI's molecular interaction format–a community standard for the representation of protein interaction data*, in "Nat. Biotechnol.", Feb. 2004, vol. 22, n^O 2, p. 177-83.
- [39] H. HERMJAKOB, L. MONTECCHI-PALAZZI, C. LEWINGTON, S. MUDALI, S. KERRIEN, S. ORCHARD, M. VINGRON, B. ROECHERT, P. ROEPSTORFF, A. VALENCIA, H. MARGALIT, J. ARMSTRONG, A. BAIROCH, G. CESARENI, D. J. SHERMAN, R. APWEILER. *IntAct: an open source molecular interaction database*, in "Nucleic Acids Res.", Jan. 2004, vol. 32, p. D452-5.
- [40] G. JEAN, D. J. SHERMAN, M. NIKOLSKI. Mining the semantics of genome super-blocks to infer ancestral architectures, in "Journal of Computational Biology", 2009, http://hal.inria.fr/inria-00414692/en/.
- [41] M. KELLIS, B. BIRREN, E. LANDER. *Proof and evolutionary analysis of ancient genome duplication in the yeast Saccharomyces cerevisiae*, in "Nature", 2004, vol. 428, p. 617-24.
- [42] M. KELLIS, N. PATTERSON, M. ENDRIZZI, B. BIRREN, E. LANDER. Sequencing and comparison of yeast species to identify genes and regulatory elements, in "Nature", 2003, vol. 423, p. 241–254.
- [43] S. KERRIEN, S. ORCHARD, L. MONTECCHI-PALAZZI, B. ARANDA, A. QUINN, N. VINOD, G. BADER, I. XENARIOS, J. WOJCIK, D. J. SHERMAN, M. TYERS, J. SALAMA, S. MOORE, A. CEOL, A. CHATR-ARYAMONTRI, M. OESTERHELD, V. STUMPFLEN, L. SALWINSKI, J. NEROTHIN, E. CERAMI, M. CUSICK, M. VIDAL, M. GILSON, J. ARMSTRONG, P. WOOLLARD, C. HOGUE, D. EISENBERG, G. CESARENI, R. APWEILER, H. HERMJAKOB. Broadening the Horizon - Level 2.5 of the HUPO-PSI Format for Molecular, in "BMC Biology", 10 2007, vol. 5, 9;5(1):44, http://hal.archives-ouvertes.fr/hal-00306554/en/.
- [44] R. KOSZUL, S. CABURET, B. DUJON, G. FISCHER. Eucaryotic genome evolution through the spontaneous duplication of large chromosomal segments, in "EMBO Journal", 2004, vol. 23, n^o 1, p. 234-43.
- [45] N. LOIRA, T. DULERMO, M. NIKOLSKI, J.-M. NICAUD, D. J. SHERMAN. Genome-scale Metabolic Reconstruction of the Eukaryote Cell Factory Yarrowia Lipolytica, in "11th International Conference on Systems Biology (ICSB 2010)", United Kingdom Edimbourg, 10 2010, P02.602, Poster, http://hal.inria.fr/ hal-00652922/en.
- [46] N. LOIRA, D. J. SHERMAN, P. DURRENS. Reconstruction and Validation of the genome-scale metabolic model of Yarrowia lipolytica iNL705, in "Journée Ouvertes Biologie Informatique Mathématiques, JOBIM 2010", France Montpellier, 09 2010, http://www.jobim2010.fr/?q=fr/node/55.

- [47] P. MARULLO, C. MANSOUR, M. DUFOUR, W. ALBERTIN, D. SICARD, M. BELY, D. DUBOURDIEU. Genetic improvement of thermo-tolerance in wine Saccharomyces cerevisiae strains by a backcross approach, in "FEMS Yeast Res", 12 2009, vol. 9, n^o 8, p. 1148–60.
- [48] P. MARULLO, G. YVERT, M. BELY, I. MASNEUF-POMARÈDE, P. DURRENS, M. AIGLE. Single QTL mapping and nucleotide-level resolution of a physiologic trait in wine Sacchar omyces cerevisiae strains, in "FEMS Yeast Res.", 2007, vol. 7, n^o 6, p. 941–52.
- [49] I. MASNEUF-POMARÈDE, C. LEJEUNE, P. DURRENS, M. LOLLIER, M. AIGLE, D. DUBOURDIEU. Molecular typing of wine yeast strains Saccharomyces uvarum using microsatellite ma rkers, in "Syst. Appl. Microbiol.", 2007, vol. 30, n^o 1, p. 75–82.
- [50] D. J. SHERMAN, P. DURRENS, E. BEYNE, M. NIKOLSKI, J.-L. SOUCIET. Génolevures: comparative genomics and molecular evolution of hemiascomycetous yeasts., in "Nucleic Acids Research (NAR)", 2004, vol. 32, p. D315-8, GDR CNRS 2354 "Génolevures" [DOI: 10.1093/NAR/GKH091], http://hal.inria.fr/inria-00407519/en/.
- [51] D. J. SHERMAN, P. DURRENS, F. IRAGNE, E. BEYNE, M. NIKOLSKI, J.-L. SOUCIET. Genolevures complete genomes provide data and tools for comparative genomics of hemiascomycetous yeasts., in "Nucleic Acids Res", 01 2006, vol. 34, n^o Database issue, p. D432-5 [DOI : 10.1093/NAR/GKJ160], http://hal.archivesouvertes.fr/hal-00118142/en/.
- [52] D. J. SHERMAN, N. GOLENETSKAYA. Databases and Ontologies for Affinity Binders, 05 2010, Overview of advances in defining ontologies and building knowledge bases for affinity binders, over the four years of the ProteomeBinders project. Presented at the Affinomics/ProteomeBinders workshop at the Møller Center, Churchill College, Cambridge University., http://hal.inria.fr/inria-00563531/en/.
- [53] H. SOUEIDAN, M. NIKOLSKI, G. SUTRE. Qualitative Transition Systems for the Abstraction and Comparison of Transient Behavior in Parametrized Dynamic Models, in "Computational Methods in Systems Biology (CMSB'09)", Italie Bologna, Springer Verlag, 2009, vol. 5688, p. 313–327, http://hal.archives-ouvertes.fr/ hal-00408909/en/.
- [54] H. SOUEIDAN, D. J. SHERMAN, M. NIKOLSKI. BioRica: A multi model description and simulation system, in "FOSBE", Allemagne, 2007, p. 279-287, http://hal.archives-ouvertes.fr/hal-00306550/en/.
- [55] L. D. STEIN. *The Generic Genome Browser: A building block for a model organism system database*, in "Genome Res.", 2002, vol. 12, p. 1599-1610.
- [56] N. VYAHHI, A. GOËFFON, D. J. SHERMAN, M. NIKOLSKI. Swarming Along the Evolutionary Branches Sheds Light on Genome Rearrangement Scenarios, in "ACM SIGEVO Conference on Genetic and evolutionary computation", F. ROTHLAUF (editor), ACM, 2009, http://hal.inria.fr/inria-00407508/en/.